Triterpenes and Saponins from Ilex argentina Leaves *

Margareth L. ATHAYDE ¹, Eloir P. SCHENKEL ^{1**}, Simone B. GNOATTO ¹, Grace GOSMANN ¹, Gustavo C. GIBERTI ² & Dominique GUILLAUME ³

¹ Faculdade de Farmácia, UFRGS, Av. Ipiranga, 2752, 90610-000 Porto Alegre, RS, Brazil. ²Centro de Estudios Farmacológicos y Botánicos, Serrano 665, 1414 Buenos Aires, Argentina. ³ Laboratoire de chimie thérapeutique, 1 rue des Louvels, 80000 Amiens, France

SUMMARY. From *Ilex argentina* Lillo leaves, a species reported as substitute or adulterant of the genuine erva-maté (*Ilex paraguariensis* A. St.-Hil.), two saponins and one triterpene have been isolated. By means of spectroscopic methods, this latter was identified as rotundic acid and the saponins as the 28-O- β -D-glucopyranosylester of rotundic acid (pedunculoside) and the 20(S) isomer of 28-O- β -D-glucopyranosylester of rotundioic acid.

RESUMEN. "Triterpenos y saponinas de las hojas de *Ilex argentina* Lillo". *Ilex argentina* es una de las especies que ha sido mencionada como adulterante o sustituto de la yerba mate verdadera (*I. paraguariensis*). De las hojas fueron aisladas dos saponinas y un triterpeno y sus estructuras químicas elucidadas a través de métodos espectroscópicos. El triterpeno fue identificado como el ácido rotúndico y las saponinas como el éster 28-O-β-D-glucopiranósido del ácido rotúndico (pedunculósido) y el isómero 20(*S*) del éster 28-O-β-D-glucopiranósido del ácido rotundioico.

INTRODUCTION

Ilex argentina Lillo is one of the species reported as adulterant or substitute of the genuine maté 1 (Ilex paraguariensis A. St.-Hil.). I. argentina's vegetative morphology is alike the one of the true erva-mate tree, so that it can be easily confused with I. paraguariensis, and it also receives one of its common names -"palo-deyerba"- due to such resemblance. Historically, a Jesuit priest seems to have been the first to note the occurrence of "yerba mate" (very probable Ilex argentina) at the subtropical rainforests close to Tucumán, Northwestern Argentina, about 1750. He also reported that such Tucumán's trees yielded a "yerba mate" not only with different taste that the one coming from Paraguay, but that it also caused headache 2. Additionally, in 1844, the governor of the Salta province authorized wild "yerba mate" harvest

in this territory for an eight year period 3. However, no one in those days was able to differentiate Ilex argentina's products from the genuine yerba mate, in spite that the problem of the bad quality of Northwestern Argentinian "yerba mate" seems to have already arisen. A few years later, Moussy 4 (1860) reported the occurrence of "Ilex mate" (botanically a synonym of Ilex paraguariensis A. St.-Hil.) for the San Francisco and Orán valleys, Salta (Northwestern Argentina). Actually, there is little doubt that Ilex mate was in fact I. argentina. Since 1936, Argentinian sanitary legislation forbids the industrialization of adulterants, among them alternative Ilex species, for the preparation of genuine verba mate.

During the course of a large program aimed to identify the adulteration of maté by other Ilex species, the systematic identification of the

KEY WORDS: Aquifoliaceae; Ilex argentina, Pedunculoside, Rotundio acid, Rotundioic acid, Saponins PALABRAS CLAVE: Acido rotúndico, Acido rotundioico, Aquifoliaceae, Ilex argentina, Pedunculósido, Saponinas

- * This work is part of the doctoral thesis of M.L.A.- PPG Ciências Farmacêuticas, UFRGS
- ** Author to whom correspondence should be addressed

triterpenoid content of the South-Brazilian *Ilex* species was undertaken ⁵⁻¹⁵. In a previous paper we have reported the isolation of the main saponin present in the leaves of *I. Argentina* ¹⁶. We report herein the isolation and structural elucidation of a novel saponin together with a known one and an acid triterpene from the same species.

MATERIAL AND METHODS General experimental procedures

SIMS and CIMS spectra were performed on a Kratos MS-80 RF spectrometer. Optical rotations were determined on a Perkin-Elmer 341 polarimeter. ¹H- and ¹³C-NMR spectra were obtained on a Bruker AC 300. Analytical TLC was carried out on silica gel Merck G and GF 254 nm plates, using chloroform:ethanol (9:1, v/v), chloroform:ethanol (8:2, v/v) and chloroform: ethanol:water (6:4:0.5, v/v) as eluant for compounds 1, 2 and 3, respectively. Detection was performed with anisaldehyde-H₂SO₄/100 °C. For TLC different authentic samples of triterpenes were used.

Plant material

Leaves of *Ilex argentina* Lillo were collected in Yerba Buena, Province of Tucumán, Argentina, in October 1992. A herbarium specimen (leg. Giberti 383) is deposited in BACP (Herbarium Cefaprin, Buenos Aires, Argentina).

Extraction and isolation

Air-dried leaves (100 g) were crushed and extracted with ethanol-water (6:4, v/v) at room temperature for seven days. The ethanol was removed under reduced pressure, the residue suspended in water and successively extracted with chloroform and ethyl acetate. Each extract was evaporated to dryness yielding 3.94 g and 2.0 g, respectively. Part of the chloroform fraction (1.0 g) was repeatedly chromatographed on a silica gel column using chloroform:ethanol (9:1 and 8:2, v/v) to afford compound 1 (13.5 mg). Ethyl acetate fraction (0.5 g) was repeatedly chromatographed on a silica gel column using chloroform:ethanol (8:2 and 7:3 v/v) resulting in 37.1 mg of pure compound 2. Another part of the ethyl acetate fraction (1.5 g) was passed through an Amberlite XAD-2 column and eluted with EtOH to give a saponin mixture fraction. Repeated separation of this mixture on a silica gel column using chloroform:ethanol (9:1, v/v) and chloroform:ethanol:water (8:4:0.5, 6:4:0.5, and 4:4:0.5, v/v) furnished 21.7 mg of compound 3.

Compound 1 and Compound 2

Identified by direct comparison with the NMR data described previously 7,14.

Compound 3

Amorphous powder, $\{\alpha\}_D = +31.85 \circ (c=0.135, \text{ MeOH}, 20 \circ \text{C})$. SIMS m/z 687 (M + Na)⁺. ¹H-NMR (C₅D₅N) δ : 1.05, 1.25, 1,25, 1.41, 1.69, (3H each, s, CH₃ x 5), 1.0 (3H, d, J = 7,0, 30-CH₃), 3.2 (1H, s, H-18) 4,7 (1H, dd, J = 6,6 and 8,5 Hz, H-3), 5,2 (1H s, 19-OH) 5.6 (1H, brt, H-12), 6.35 (1H, d, J = 8.0 Hz, Glc-1). ¹³C-NMR (C₅D₅N): (C1 to C30)39,0; 27,8; 75,5; 54,4; 51,9; 21,8; 33,3; 40,8; 48,0; 36,8; 24,0; 127,4; 138,8; 42,1; 29,1; 26,7; 48,2; 47,2; 73,3; 42,8; 24,6; 31,8; 180,7; 12,9; 16,1; 17,4; 24,2; 176,9; 29,6; 16,0; glucopyranosyl moiety (C1 to C6) 95,8; 74,0; 78,9; 71,0; 79,3 and 62,1.

RESULTS AND DISCUSSION

The alcoholic extract of the leaves of *I. argentina* was concentrated and successively extracted with solvents of increasing polarity (chloroform and ethyl acetate). Compound **1** (13.5 mg) was isolated from the chloroform fraction. Its ¹³C-NMR spectrum revealed the presence of 30 carbons indicating its triterpenic nature and the lack of sugar moiety. Careful analysis of the ¹³C-NMR of **1** and co-TLC revealed that it was rotundic acid, a triterpene that we had previously isolated from *I. taubertiana* ⁹ and from *I. theezans* ¹⁴.

Compound **2** (37.1 mg) was isolated from the ethyl acetate fraction. Its 1 H- and 13 C-NMR data indicated its triterpene nature (36 carbons). The hexose could be identified as one β -glucosyl moiety (δ 6.29, 1H, d, J = 8 Hz, H-1'; δ 95.8, C-1'), both chemical shifts evidencing an ester linkage between the sugar portion and the aglycone. A careful comparison of the 13 C-NMR data obtained for **2** with the literature indicated that **2** was pedunculoside (Figure 1) a triterpenoid previously isolated from *I. rotunda, I. pedunculosa* and *I. oldhami* 17,18 and that we had previously identified from *I. taubertiana* 7 and *I. theezans* 14 .

Compound **3** (21.7 mg) was isolated from the ethyl acetate fraction. Its 1 H-NMR indicated the presence of the β -glucosyl unit (H-1: δ 6.35, d, J = 8 Hz) linked to the aglycone by an ester linkage. Similarly to **2**, its 13 C-NMR spectrum displayed signals for 36 carbons, but the major differences between the two spectra include characteristic signals due to one additional carboxyl group in **3** (δ 180.7) instead of an hydroxymethylene group in **2**. Comparing the chemical

Figure 1. Compounds 1, 2 and 3 isolated from *Ilex argentina* leaves.

shifts assigned in the A ring of 3 with those of 2, we observed that the C4, C5 and C6 shifted from δ 42.8 to 54.4, 48.5 to 51.9 and 18.7 to 21.8 ppm respectively. These data are coherent with those observed ¹⁹ when C23-αCH3 is replaced with COOH that resembled those of the 28-O-β-D-glucopyranosylester of rotundioic acid that we had already isolated from I. theezans 14. However, NMR comparison indicated that the two compounds, despite close similarities in A ring, were particularly different on E ring, suggesting another stereochemistry at C20. NMR data to discriminate between 20(S) and 20(R) series are numerous 7,20-22, and the striking differences concern the chemical shifts of C18, C22 and C29. Since we had in hand both isomer, we could observe that indeed, in the case of 3, C18' was at 47.2 ppm and C22 at 31.8 ppm, whereas we observed these signals at 52.8 and 36.2 ppm respectively in the case of the acetylated 28-Oβ-D-glucopyranosylester of rotundioic acid ¹⁴. Also, the chemical shift observed for C29 was shifted by ca. 2.9 ppm downfield (29.6 ppm), when compared with those reported for the 20(R) series. Thus 3 was unambiguously identified as the 20(S) isomer of $28-O-\beta-D$ -glucopyranosylester of rotundioic acid (Figure 1). To the best of our knowledge, this is the first report of the natural occurrence of this compound. Very few 19α-hydroxyursolic acid derivatives with the singular 20S-configuration have been isolated, most of them from *Ilex species* 7,15,20-23.

Considering the exomorphological resemblance, I. argentina has been provisionally classified in the same taxonomic group to which I. paraguariensis belongs, according to Loesener ²⁴, *i.e.*: subsection *Repandae*, section *Microdon*tae, series Aquifolium, subgenus Euilex. However, phytochemical analysis indicates a very different chemical composition, not only regarding the absence of xanthines or presence in

traces 25,26, but also as a result of the present study on saponins. Both species, or their extracts, can be easily distinguished by the chromatographic profile of the saponins constituents, as shown in Figure 2, in which the substances were detected by spraying with anisaldehyde in sulfuric acid and heating, enabling maximal color development and visualisation in visible and long-wavelength UV light. In com-

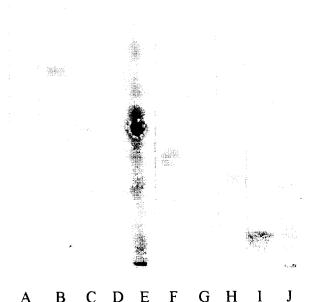


Figure 2. TLC profile of Ilex argentina and Ilex paraguariensis. Eluent: chloroform:ethanol:water (60:40:5 by volume), detection with anysaldehyde-sulfuric acid followed by heating . A: compound 1 (rotundic acid); B: compound 2 (pedunculoside); C: ILA₁ solated from I. argentina 16; D. compound 3; E. n-butanol fraction of I. argentina leaves; F: isoquercitrin; G: rutin; H: matessaponin 1 (isolated from I. paraguariensis leaves); I: matessaponin 2 (isolated from I. paraguariensis leaves 5,9); J: n-butanol fraction of I. paraguariensis leaves 10.

В

parison with the glycosides isolated from I. paraguariensis which are derivatives of ursolic and oleanolic acids (deep purple color under visible and a bright yellow under UV light), the glycosides isolated from I. argentina (19 α -hydroxyursolic acid derivatives) displayed a blue color under visible light and a pale yellow color under UV conditions. These features could be helpful to allow the differentiation between the two species from the chemical point of view. Additionally, both chromosome numbers and DNA content were found to be different in both

species 27 -for *I. paraguariensis* n = 20 and DNA content 2C = 2.23 ± 0.08 pg; whilst for *I. argentina* n = 40 and DNA content 2C = 4.27 ± 0.07 pg.

Acknowledgments. We are grateful to Prof. Dr. Nilo Zanatta - Departamento de Quimica- Universidade Federal de Santa Maria, for NMR spectroscopic measurements of compounds **1** and **2**. This work was supported by fellowships from PICDT-CAPES (Brazil) for M. L. Athayde and CNPp research fellowships for G. Gosmann and E.P. Schenkel.

REFERENCES

- 1. Giberti, G.C. (1989) Dominguezia 7: 3-21
- Grondona, E.M. (1953) Revista Argent. Agron.
 68-95
- Anonymous (1930) La Industria azucarera 35 (436): 106-8
- 4. Moussy, V.M. de (1860) Libraire Firmi Didot Frères, Fils et Cie., Paris: pp. 582
- Gosmann, G., E.P. Schenkel & O. Seligmann (1989) J. Nat. Prod. 52: 1367-70
- 6. Taketa, A.T.C. & E.P. Schenkel (1994) Acta Farm. Bonaerense 13: 159-64
- 7. Taketa, A.T.C. & E.P. Schenkel (1995) *Rev. Bras. Farm.* **76**: 9-11
- 8. Heinzmann, B.M. & E.P. Schenkel (1995) *J. Nat. Prod.* **58**: 1419-22
- Gosmann, G., D. Guillaume, A.T.C. Taketa & E.P. Schenkel (1995) J. Nat. Prod. 58: 438-41
- Schenkel, E.P., J.A. Montanha & G. Gosmann (1996) "Triterpene saponins from maté, *Ilex paraguariensis*", en "Saponins used in food and agriculture" (G.R. Waller & K. Yamasaki, eds.), Plenum Press, New York, pp. 47-56
- Kraemer, K.H., A.T.C. Taketa, E.P. Schenkel, G. Gosmann & D. Guillaume (1996) Phytochemistry 42: 1119-22
- Pires, V.S., D. Guillaume, G. Gosmann & E.P. Schenkel (1997) J. Agric. Food Chem. 45: 1027-31
- Schenkel, E.P., G. Gosmann, J.A. Montanha, B.M. Heizmann, M.L. Athayde, A.T.C. Taketa, V.S. Pires & D. Guillaume (1997) Ciência e Cultura 49: 359-63
- 14. Athayde, M.L., E.P. Schenkel, G. Gosmann & D. Guillaume (1999) *Acta Farm. Bonaerense* **18**: 49-52

- Taketa, A.T.C., T. Schmittmann-Schlager, D. Guillaume, G. Gosmann & E.P. Schenkel (2000) Phytochemistry 53: 901-04
- Schenkel, E.P., M.L Athayde, G.C. Giberti & D. Guillaume (1995) Acta Farm. Bonaerense 14: 5-10
- Nakatani, M., Y. Miyazaki, T. Iwashita, H. Naoki & T. Hase (1989) Phytochemistry 28: 1479-82
- 18. Amimoto, K., K. Yoshikawa & S. Arihara (1992) Chem. Pharm. Bull. 40: 3138-41
- Tan, N., J. Zhou & S. Zhao (1999) Phytochemistry 52: 153-92
- 20. Kakuno, T., K. Yoshikawa & S. Arihara (1992) *Phytochemistry* **31**: 3553-7
- 21. Ouyang, M., H. Wang, Y. Liu & C. Yang (1997) Phytochemistry 45: 1501-5
- 22. Ouyang, M., Y. Liu, H. Wang & C. Yang (1998) Phytochemistry **49**: 2483-6
- Hidaka, K., M. Ito, Y. Matsuda, H. Kohda, K. Yamasaki, J. Yamahara, T. Chisaka, Y. Kawakami, T. Sato & K. Kagei (1987) Chem. Pharm. Bull. 35: 524-9
- Loesener, T. (1901) Monographia Aquifoliacearum. I. Nova Acta Acad. Caes. Leop. - Carol. German. Nat. Cur. 78: pp. VIII + 600 + 15 tabs.
- Filip, R., D.I.A. Iglesias, R.V.D. Rondina & J.D. Coussio (1983) Acta. Farm. Bonaerense 2: 87-90
- Filip, R., G.E. Ferraro, R.V.D. Rondina & J.D. Coussio (1989) An. Asoc. Quim. Arg. 77: 293-7
- Barral, G.; L. Poggio & G.C. Giberti (1995) Bol. Soc. Argent. Bot. 30 (3-4): 243-8